

Amendments to the Specification:

Please replace the paragraph beginning at page 10, line 10 with the following rewritten paragraph:

RNA was reverse-transcribed with Moloney murine leukemia virus reverse transcriptase (Perkin-Elmer). RNA samples (0.5 µg of total RNA), 0.5 unit of RNase inhibitor, 1 mM of each dNTP, 0.5 nmol/ml of specific 3' primer, 5 mM MgCl₂ and 1.25 units of reverse transcriptase were incubated in a total volume of 10 µl of reaction mixture containing the enzyme buffer supplied by the manufacturer. The reaction mixture was incubated for 30 min at 42°C, and then heated for 5 min at 95°C. The reverse-transcribed products were then amplified with Gold DNA polymerase (Perkin Elmer) on a thermocycler (Biolabo, Scientific Instruments, Chatel St Denis, Switzerland). The polymerase chain reaction (PCR) was performed in a total volume of 50 µl using 10 µl of the reverse-transcribed products in PCR buffer, 2 mM MgCl₂, 5 µM of each dNTP, 0.2 nmol/ml of both OPG-specific antisense

ACTAGTTATAAGCAGCTTATTTTACTG (SEQ ID NO: 2),

and sense

GGAGGCATTCTTCAGGTTTGCTG (SEQ ID NO: 3)

primers and 1.25 units of DNA polymerase. After an initial denaturation step of 10 min at 95°C, samples were amplified by 35 cycles of denaturation at 94°C for 45 sec, annealing at 60°C for 1 min, and extension at 72°C of 1 min 30 sec, followed by a 7-min extension step at 72°C. All samples were subjected to RT-PCR with 13-actin as a positive control. Samples of RT-PCR products were loaded onto a 1.2 % agarose gels (containing ethidium bromide) in 1 x T AE buffer and separated by electrophoresis at 150 V for 1 hr. RT-PCR products were visualised under UV light. The correct size of the bands was determined by comparison with DNA size markers (Boehringer Mannheim).

Please replace the paragraph beginning at page 13, line 1 with the following rewritten paragraph:

With the OPG specific antisense primer

CCGGCCTCTTCGGCCGCCAAGCGAGAAACGTTTCCTCCAAAGTACC

(SEQ ID NO: 4), and the sense primer

ACTAGTTATAAGCAGCTTATTTTACTG (SEQ ID NO: 5),

a 1174 bp PCR fragment was amplified from this cDNA. The PCR product was *Sfi*I-*Spe*I digested, gel purified and the resulting 1156 bp fragment ligated to *Sfi*I XbaI digested and SAP-treated pINA1267, creating pNFF270. This plasmid pNFF270 was then introduced into the yeast *Yarrowia lipolytica* by transformation. Figure 3 depicts the restriction map of the plasmid which was integrated into the genomic DNA of *Yarrowia* transformants and SEQ ID. No. 1 the protein encoded by this OPG plasmid.